

# Fluorescein-based isoelectric point markers as a tool for tracking of pH gradient in highly sensitive isoelectric focusing analysis with laser induced fluorescence detection

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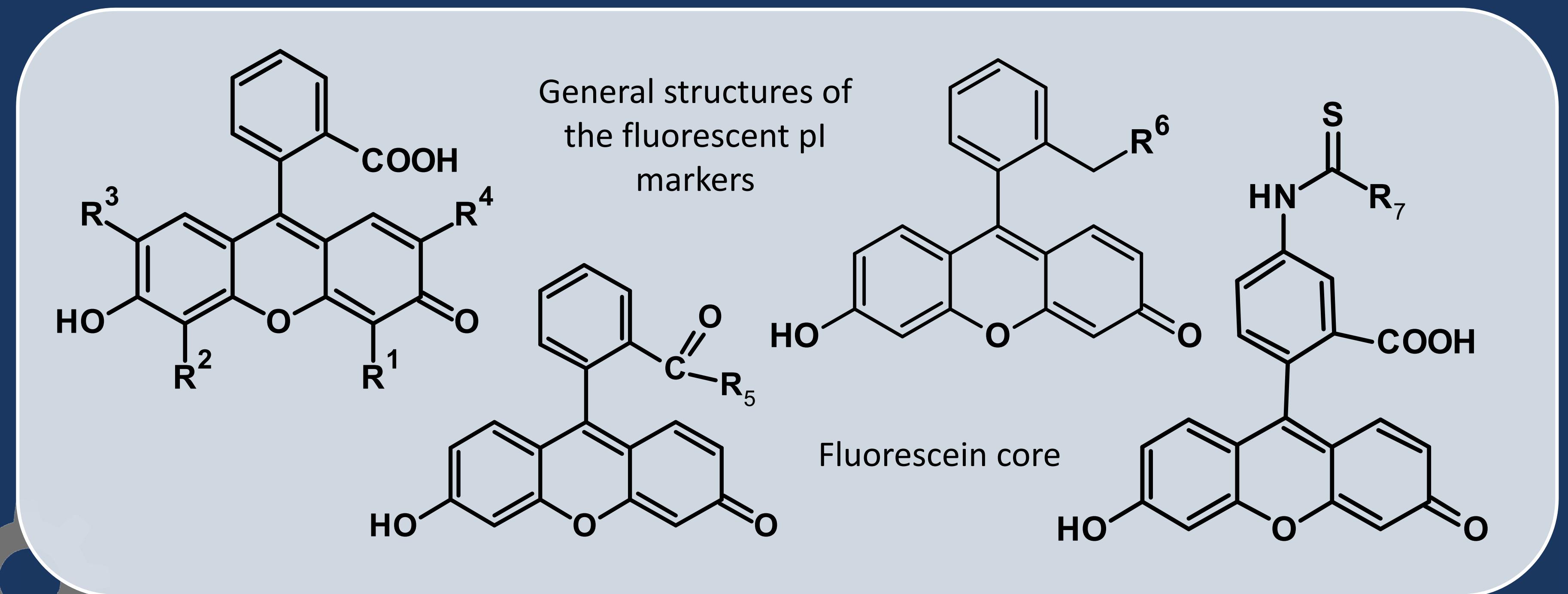
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Pavlína Dadajová<sup>1,2</sup>, Filip Duša<sup>1</sup>, Richard Čmelík<sup>1</sup>, Karel Šlais<sup>1</sup>

<sup>1</sup>Institute of Analytical Chemistry of the Czech Academy of Sciences, Czech Republic

<sup>2</sup>Department of Chemistry, Faculty of Science, Masaryk University, Czech Republic

Isoelectric point (p<sub>l</sub>) markers are crucial internal standards used in isoelectric focusing (IEF) methods for tracking the pH gradient. Our laboratory has already introduced colored (UV and visible region absorbing) low-molecular-mass (LMM) p<sub>l</sub> markers for capillary isoelectric focusing (cIEF) as an alternative to the commonly used peptide and protein p<sub>l</sub> markers. The cIEF instrumentation can be coupled with on-line laser induced fluorescence (LIF) detection, which significantly increases detection sensitivity. However, there was considerable lack of suitable p<sub>l</sub> markers for this type of application. Consequently, our laboratory devised fluorescein-based low-molecular p<sub>l</sub> markers, initially covering narrow pH range (from 5.4 to 6.6). By adding various functional groups to the fluorescein core. Of all the synthesized and tested candidate structures 21 markers were identified as suitable for the cIEF-LIF method.

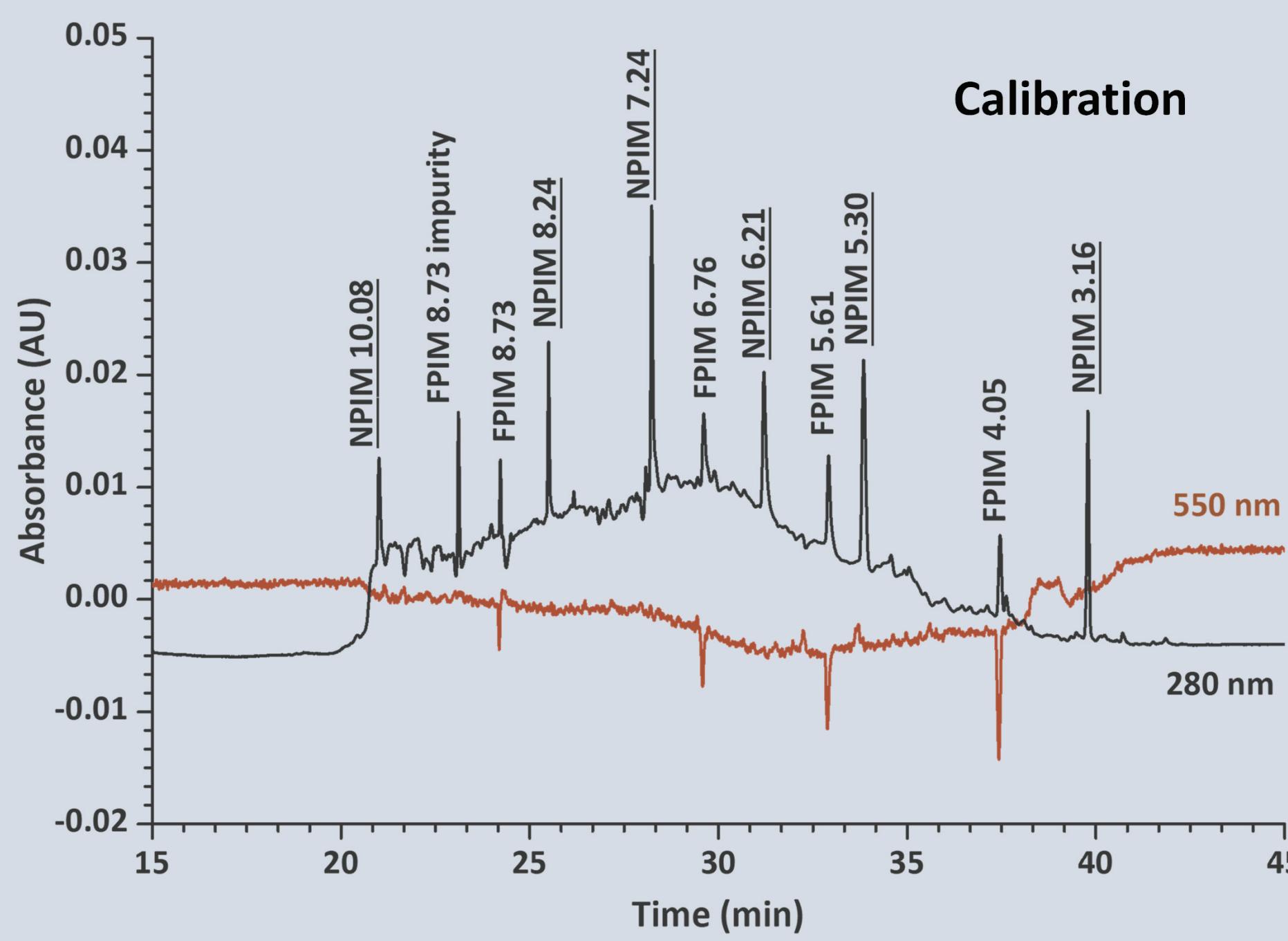


**Instrument:** Sciex P/ACE™ MDQ Plus, DAD (280 nm) and LIF (ex 488/em bandpass 520/20 nm and longpass 550 nm), neutral LPA capillary 30.2/20 cm, cooling at 20°C

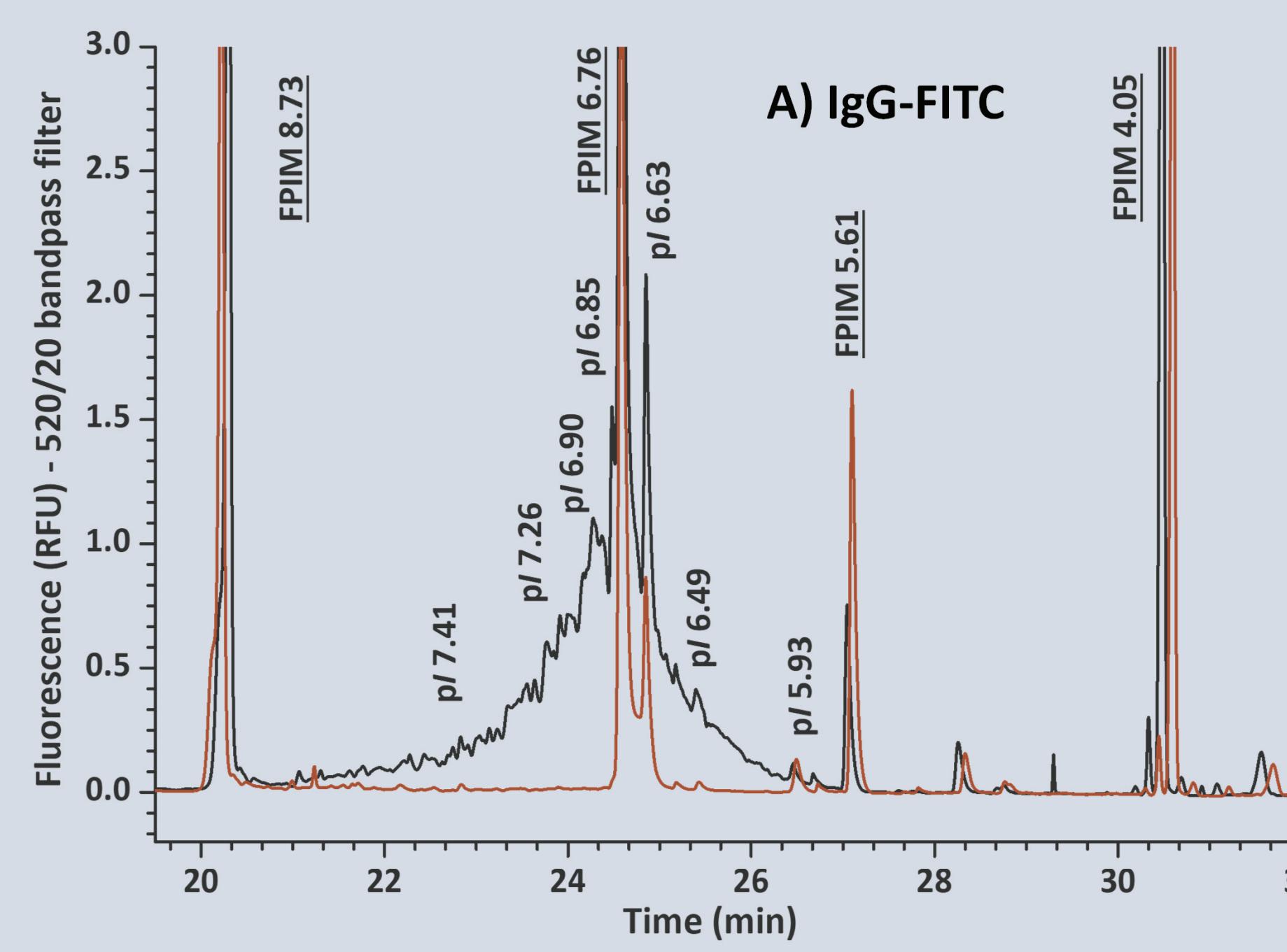
**Separation conditions:** Two step cIEF – focusing 200 mM H<sub>3</sub>PO<sub>4</sub> and 300 mM NaOH (30 kV, 15 min), chemical mobilization with 350 mM CH<sub>3</sub>COOH (25kV).

**Sample:** 2.4 M urea, 80% V/V cIEF gel, 1.92% w/V background ampholytes (Pharmalyte 3-10), 60 mM L-arginine and 1.6 mM iminodiacetic acid

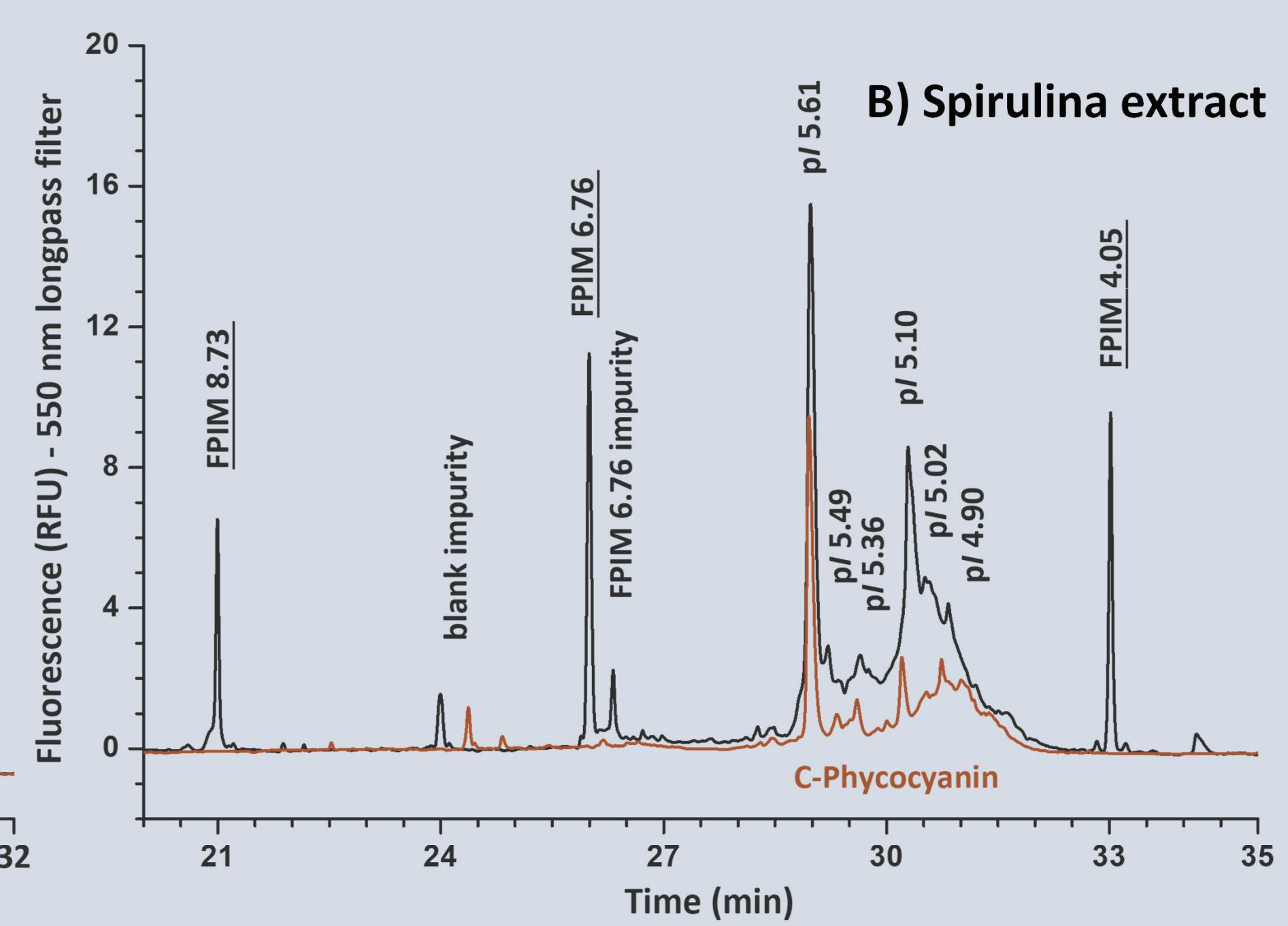
Label Main peak p <sub>l</sub>	SD n=3	Notable impurities at p <sub>l</sub> :	Purity (%)
<b>FPIM 3.10</b>	0.001	3.35	63
<b>FPIM 4.05*</b>	0.002	-	87
<b>FPIM 4.06</b>	0.004	-	90
<b>FPIM 4.07</b>	0.0003	3.73	67
<b>FPIM 5.31</b>	0.003	6.47	39
<b>FPIM 5.44</b>	0.0001	-	80
<b>FPIM 5.61*</b>	0.002	-	97
<b>FPIM 5.88</b>	0.001	-	82
<b>FPIM 6.40</b>	0.002	5.0–6.4	67
<b>FPIM 6.45</b>	0.002	6.0–6.3	82
<b>FPIM 6.47</b>	0.001	-	97
<b>FPIM 6.62</b>	0.001	5.61	67
<b>FPIM 6.72</b>	0.003	-	80
<b>FPIM 6.76*</b>	0.001	-	92
<b>FPIM 6.77</b>	0.001	-	92
<b>FPIM 6.78</b>	0.0004	-	95
<b>FPIM 7.03</b>	0.001	-	95
<b>FPIM 8.27</b>	0.001	7.8–8.3	60
<b>FPIM 8.73*</b>	0.002	9.85	74
<b>FPIM 9.25</b>	0.001	6.0–6.1, 9.2–10.0	40
<b>FPIM 10.21</b>	0.008	9.4–10.2	45



► Purity assessment and calibration of four selected fluorescent markers (marked by \* in the table) in UV 280 nm with our yellow nitrophenol p<sub>l</sub> markers. Calibration of the remaining markers was with LIF detection.



► Application of the markers in p<sub>l</sub> determination and characterization of A) commercially labeled antibody - IgG-FITC ( $\lambda_{\text{max}} \sim 512$  nm), where multiple labeled isoforms of the antibody can be observed and B) extract from *Arthrospira platensis* (Spirulina) with comparison to a C-Phycocyanin standard ( $\lambda_{\text{max}} \sim 640$  nm), which is at its p<sub>l</sub> present in a monomer-trimer system.



## CONCLUSION

We present 21 new characterized fluorescent ampholytic structures for tracking of the pH gradient. Their p<sub>l</sub> values have been determined with precision of one hundred of pH unit using a linear calibration based on four selected markers. The selected structures have been proved to be p<sub>l</sub> markers featuring good focusing abilities, manifested as well-defined narrow peaks that can be easily applied in IEF methods with fluorescence detection and cover the range from 3.10 to 10.21, which greatly expands the selection of fluorescent p<sub>l</sub> markers.

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